C. cerastes were capable of feeding on White-tailed Antelope Squirrels (*A. leucurus*). Here, we document an instance of a *C. cerastes* attempting to ingest an *A. leucurus*, which resulted in the death of both animals.

On 2 April 2001, one of us (BC) found a pair of courting *C. cerastes* on the eastern edge of the Kelso Dunes in Mojave National Preserve, San Bernadino County, California, USA. The pair was captured and the female (567 mm SVL; 133 g) was implanted with a radio-transmitter, released, and tracked until 31 May 2001 (the end of the spring field season). During this period she moved an average of 4.8 m/day. On 28 August 2001 (the beginning of the autumn field season), the signal was again located. The snake was 157.4 m from her 31 May location and was below ground in a dune stabilized by Big Galleta Grass (*Pleuraphis rigida*). The signal remained stationary throughout the entire autumn and following spring field seasons. We excavated the dune on 21 May 2002 and discovered the mummified body of the snake with the remains of an *A. leucurus* in her mouth. The pair was lying on their backs in a burrow system about 20 cm from the nearest entrance. The head of the snake was twisted slightly to her left and her mouth held the antelope squirrel around the shoulder.

It is unclear exactly how the snake died, but it is possible that after engaging her teeth to begin feeding, she was constrained by the position of the squirrel in the burrow and was unable to either continue swallowing or release the prey item. In this scenario, the cause of death would either be starvation or suffocation. Alternatively, the burrow might have collapsed as a result of movements associated with feeding, resulting in suffocation. Both specimens are deposited in the Soda Springs Desert Studies Center, Zzyzx, California (*Crotalus cerastes*, SSDSC 59; *Ammospermophilus leucurus*, SSDSC 60).

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ELAPHE GUTTATA EMORYI (Great Plains Ratsnake). **DIET.** *Elaphe guttata emoryi* eats a wide variety of prey including rodents, ground nesting birds, and occasionally lizards (Werler and Dixon 2000. Texas Snakes: Identification, Distribution, and Natural History, University of Texas Press, Austin, Texas. 437 pp.). On 7 October 2006 (1100–1130 h) we collected an adult male *E. g. emoryi* (154 mm SVL, 72 mm TL, 124 g) under green briar vines in a live oak motte at Camp Bowie National Guard Training Facility (31°35'44.6"N, 098°53'58.7"W, 429 m elev.), Brown County, Texas, USA. The snake had an adult female *Sceloporus olivaceus* (103 mm SVL, 153 mm TL, 44 g) in its stomach that was consumed headfirst. The lizard and snake are deposited in the Angelo State Natural History Collection (ASNHC 14192-93), Angelo State University, San Angelo, Texas.

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MASTICOPHIS FLAGELLUM (Eastern Coachwhip). **REPRODUCTION.** The reproductive ecology of *Masticophis flagellum* in the northwestern Gulf Coastal Plain near the convergence of the borders of northeast Texas, southeast Oklahoma, southwest Arkansas, and northwest Louisiana is poorly understood. Two *M. flagellum* clutches from northern and central Arkansas contained 14 and 18 eggs (Trauth et al. 1994. Proc. Arkansas Acad. Sci. 48:196–209). Clutches from Oklahoma contained 7, 12, and 13 eggs, and were oviposited from early to mid-July (Carpenter 1958. Herpetologica 14:113–115). A single female (1160 mm SVL, 322 g) from Smith County, Texas laid a clutch of 11 eggs on 29 June 1988 (Ford et al. 1990. Texas J. Sci. 42:355–368). Eggs from that clutch were 4.47 (SD = 0.29) cm long, 2.33 (SD = 0.05) cm wide, and weighed 15.3 (SD = 9.61) g.

On 14 June 2006, a female *M. flagellum* (1154 mm SVL, 1484 mm TL, 465.2 g [mass taken prior to egg removal]) was recovered from a rubbish pile ca. 4 km from the Arkansas-Texas border along George Thomas Road (33.33098-N, 94.0852-W, elev. 129 m, Datum: NAD27) and ca. 0.75 km S of the junction of this road with Fricks Road in the Liberty Eylau region of Texarkana, Bowie County, Texas. The animal was placed in cool storage (2.5°C) until 15 June 2006 when it was removed for photography. After approximately 30 min at 30°C the animal was listless and its vent appeared dilated for oviposition. I placed the animal in a 40 L aquarium with a lid, water, and a terra cotta flower pot as a shelter. On 16 June 2006 at about 0830 h the specimen was found dead. Fourteen eggs were dissected from the abdominal cavity, weighed with an electronic balance, and measured using Vernier calipers. Mass of individual eggs (mean = 11.16 g, SE = 0.097) was not normally distributed (Anderson-Darling: $A^2 = 0.739$, $P = 0.041$), whereas length (mean = 37.88 mm, SE = 0.377; $A^2 = 0.213$, $P = 0.815$) and diameter (mean = 22.82 mm, SE = 0.213; $A^2 = 0.383$, $P = 0.347$) were normally distributed. Eggs had the following measurements based on their location in the oviduct: Cranial (N = 5): mean mass = 11.1 g (SE = 0.1), mean length = 36.5 mm (SE = 0.5), mean diameter = 22.9 mm (SE = 0.2); Middle (N = 5): mean mass = 11.3 g (SE = 0.22), mean length = 38.3 mm (SE = 0.5), mean diameter = 22.5 (SE = 0.2); Caudal (N = 4): mean mass = 11.1 g (SE = 0.2), mean length = 37.2 (SE = 1.0), mean diameter = 23.2 (SE = 0.7). Although the absolute mean dimensions of eggs located centrally in the reproductive tract seemed larger, the position of the eggs in the reproductive tract was not a significant indicator of an egg's mass (Kruskal-Wallace: $H = 1.09$, $df = 2$, $P = 0.580$), length (ANOVA: $F = 0.71$, $df = 13$, $P = 0.501$), or diameter (ANOVA: $F = 0.730$, $df = 13$, $P = 0.504$).

The timing of reproduction by this female mirrors observations in northeast Arkansas and southeast Oklahoma (Carpenter, *op. cit.*; Trauth et al., *op. cit.*) and is only two weeks earlier than observed in Smith County, Texas (Ford et al., *op. cit.*). The clutch size in this observation is identical to the observation from Smith County,